

# Antonia Red™-lysine-dextran

CAS nr: N/A

Structure:

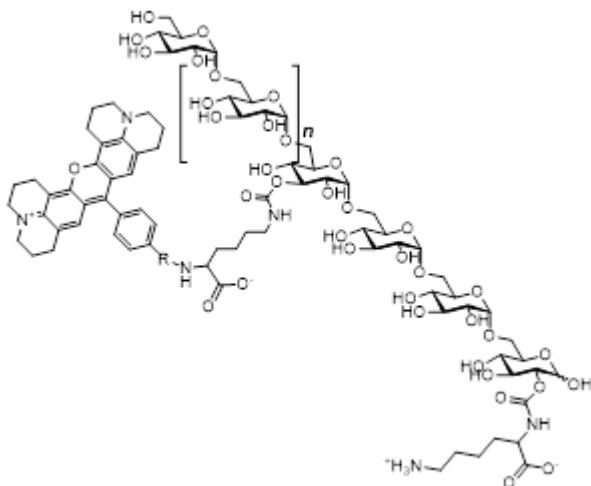


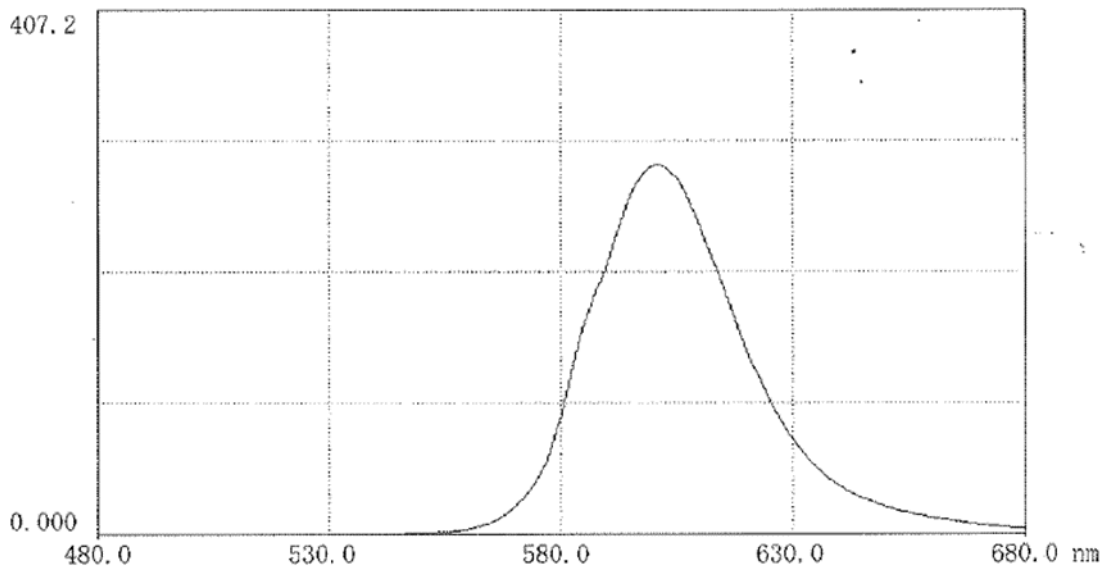
Fig. 1. Structural representation of Antonia Red-lysine-dextran.

## Synthesis and structure

Lysine Dextrans are synthesised from well-characterized dextran fractions derived from *Leuconostoc mesenteroides* labelled with Lysine. Lysine dextran is then functionalized with Antonia Red under mild conditions to form Antonia Red Lysine Dextran. After purification, the products are controlled for Mw, appearance, solubility, DS, pH, free Lysine and free dye. The products are designated by the approximate molecular weights of the dextran fractions used. Thus, for example, the product Antonia Red Lysine Dextran 4 has a molecular weight of approx. 4000 Da. The actual molecular weight is determined by GPC. This value is supplied with the Certificate of Analysis. The dextran used is from *Leuconostoc mesenteroides*\_B-512F which is essentially a linear  $\alpha$ -(1-6)-linked glucose chain with however a low percentage (2-5%) of  $\alpha$ -(1-3) branches distributed along the chain. The dextran fractions used are from Mw of 4000 to 500000 and are carefully controlled by GPC, optical rotation, absorbance and other control parameters.

## Physical properties

Antonia Red™ is a novel red-shifted dye similar to Texas Red and is developed and produced by TdB Consultancy AB. Antonia Red Lysine Dextrans are purple powders that are readily soluble in water or electrolyte solutions. Antonia Red Lysine Dextran are insoluble in most organic solvents, such as ethanol, methanol, acetone, chloroform, ethyl acetate etc. The degree of substitution, (DS) is between 0.01-0.02 (mol Lysine/mol Glucose) and 0.001-0.01 (mol Antonia Red/mol Glucose).



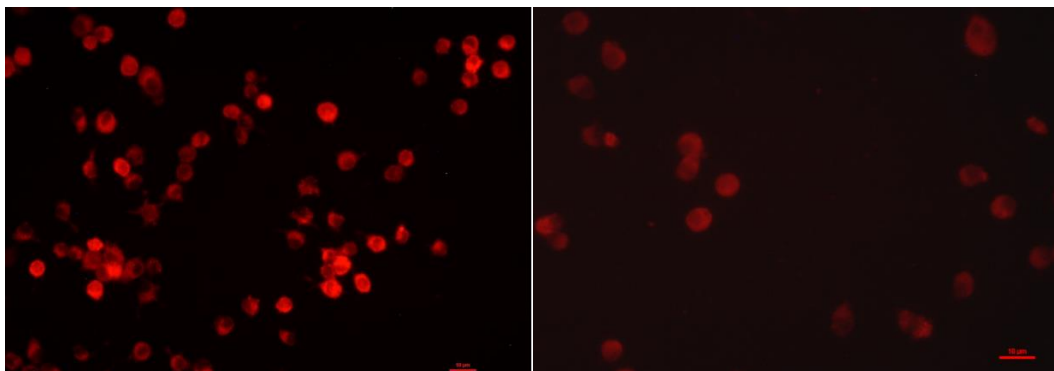
*Figure 2. Fluorescence scan of Antonia Red Lysine Dextran 70 in 0.025M Borate pH 9 buffer. Excitation 583 nm, Emission 602 nm.*

## Stability

No prospective stability studies on Antonia Red Lysine Dextrans have been performed yet. However, the structural properties of the dextran and of the carbamide linkage of the Lysine to the dextran chain as well as the covalent bond between Lysine and Antonia Red would suggest high stability of the product. It is recommended that the products are stored in air-tight containers. Antonia Red Lysine Dextrans may be stored at ambient temperatures.

## Applications

Dextrans carrying amino groups are valuable to the scientific community as versatile tools for bioconjugation and fixation in living systems<sup>1</sup>. For conjugation, the free amino group of the Lysine structure will be able to covalently bind to activated entities such as NHS-esters or isothiocyanates or be coupled to carboxylic acids using common peptide coupling reagents such as DCC, HOBT and or HATU. Cell and tissue fixation is performed in order to preserve components in a “life-like state” and to make cells permeable to allow antibodies to access cellular structures, for example for microscopy studies or immunostaining. The amino groups on Lysine Dextran reacts with the fixing agent (here: formaldehyde or glutaraldehyde) to form a covalent crosslinking with biomolecules such as proteins and lipids, immobilizing the whole system and the dextran. This is of particular importance when evaluating biological events qualitatively or quantitatively in molecular imaging. Without fixation, these structures within a living system would fall apart and diffuse rapidly. Antonia Red is a fluorescent dye that has an excitation maximum at 583 nm and an emission maximum at 602 nm, making it an analogue of the popular Texas Red dye (Figure 3).



*Figure 3. Pictures of macrophages RAW264.7 in a Nikon TE2000-U microscope demonstrating the superior signal from Antonia Red-dextran (Left) compared to Texas Red Dextran, lysine fixable (Right). Antonia Red-dextran shows superior fluorescent signal, less aggregation, less free dye and excellent cellular uptake.*

## References

1 (a) Henley JR, Krueger EW, Oswald BJ, McNiven MA, *J Cell Biol* (1998) 141:85-99; (b) Fritzsich B, Christensen MA, Nichols DH; *J Neurobiol.* 1993 Nov;24(11):1481-99; (c) Fritzsich B, *J Neurosci Methods* (1993) 50:95-103.