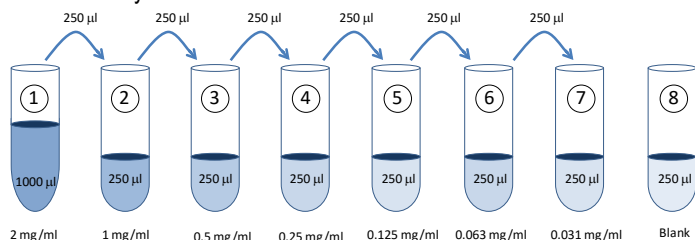




## ASSAY PROCEDURE

- Prepare Standard Dilutions:** The recommended standard range is 0.031 - 2 mg/ml. Dissolve one vial of Albumin Standard in 1 ml of PBS for the 2 mg/ml standard. Then serially dilute it with PBS. For example, mix 200  $\mu$ l of the standard (2 mg/ml) with an equal volume of PBS to make a 1 mg/ml solution, and then repeat it five more times for 0.5, 0.25, 0.125, 0.063, and 0.031 mg/ml solutions. The remaining 2 mg/ml standard stock may be stored at -20°C for use in a second assay. We recommend making fresh serial dilutions for each assay.



- Prepare Sample Dilutions:** Dilute samples with PBS. It is recommended to use 2-3 different dilutions if the sample albumin level is unknown.
- Prepare Dye Solution:** Mix 150  $\mu$ l of Solution A with 50  $\mu$ l of Solution B for each well just before use. For example, 8 samples, 7 point standards, and one blank (all in duplicate) will require 6.4 ml of the Dye Solution. Mix 4.8 ml of Solution A with 1.6 ml of Solution B.
- Add Standards and Samples:** Use the plate layout as shown in Figure 1. Add 50  $\mu$ l of standards, PBS (Blank), and samples to designated wells.
- Add Dye Solution:** Add 200  $\mu$ l of the dye solution into all wells. Incubate at room temperature for 5 minutes.
- Read Plate:** Read the OD values at 610 nm. If the OD values of samples are greater than the OD values of the highest standard, re-assay the samples at a higher dilution.

## CALCULATION OF PROTEIN CONCENTRATION

- Average the duplicate OD values for the standards, blank (B), and test samples.
- Subtract the "blank" (B) values from the averaged OD values of the standards and test samples.
- Plot the OD values of standards against the mg/ml of standard. Using a log/log plot will linearize the data. Figure 2 shows a representative experiment where the standard range is from 0.031-2 mg/ml.
- The mg/ml of protein in test samples can be calculated using regression analysis. Multiply it by the sample dilution factor to obtain the protein concentration (mg/ml) in original test samples. For additional assistance, please download a sample calculation worksheet from Chondrex, Inc.'s web site.

Figure 2 - A typical standard curve for BPB protein assay

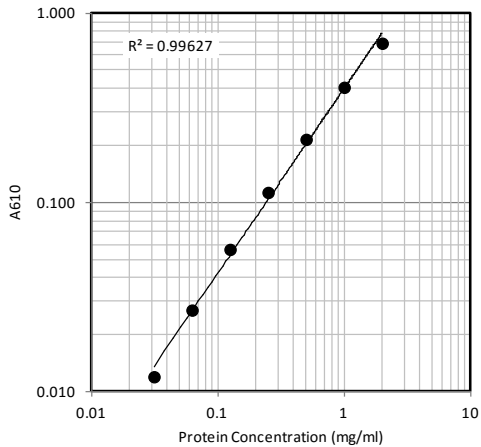


Table 1 - Reproducibility of data assayed by BPB Protein Assay Kit

Test At	1 mg/ml	0.3 mg/ml	0.07 mg/ml
Inter-Assay CV (%)	4.4	6.2	9.8
Intra-Assay CV (%)	3.3	5.2	1.7
Spiking Test*	92.5 %	100.2 %	104.3%

Standard was added with known amounts of albumin and then diluted with PBS to assay the protein concentration.

## REFERENCES

1. D. Ellis, G. J. Buffone, New approach to evaluation of proteinuric states. *Clin Chem* 23, 666-670 (1977).
2. P. A. Peterson, P. E. Evrin, I. Berggård, Differentiation of glomerular, tubular, and normal proteinuria: determinations of urinary excretion of  $\beta$ 2-microglobulin, albumin, and total protein. *J Clin Invest* 48, 1189-1198 (1969).
3. W. L. Gyure, Comparison of several methods for semiquantitative determination of urinary protein. *Clin Chem* 23, 876-879 (1977).
4. B. C. Garner, C. E. Wiedmeyer, Comparison of a semiquantitative point-of-care assay for the detection of canine microalbuminuria with routine semiquantitative methods for proteinuria. *Vet Clin Pathol* 36, 240-244 (2007).
5. L. M. Killingsworth, Clinical applications of protein determinations in biological fluids other than blood. *Clin Chem* 28, 1093-1102 (1982).
6. K. H. Schosinsky, M. Vargas, A. Luz Esquivel, M. A. Chavarria, Simple spectrophotometric determination of urinary albumin by dye-binding with use of bromophenol blue. *Clin Chem* 33, 223-226 (1987).
7. R. Flores, A rapid and reproducible assay for quantitative estimation of proteins using bromophenol blue. *Anal Biochem* 88, 605-611 (1978).
8. C. S. Greenberg, P. R. Craddock, Rapid single-step membrane protein assay. *Clin Chem* 28, 1725-1726 (1982).