Chondrex, Inc.

Cell Culture Grade Bovine Type | Collagen

Catalog # 1202

For Research Use Only - Not Human or Therapeutic Use

Collagen, the most abundant protein in vertebrates, is observed in skin, cartilage, bone, intervertebral discs, blood vessels, tendons, ligaments, and corneas, and is the main component of the extracellular matrix (ECM). Type I collagen consists of two identical alpha 1 chains and one distinct alpha 2 chain (1), forming a triple helix formation known as tropocollagen. This triple-stranded helical conformation increases structural strength and resistance to enzymatic degradation in tissue and plays a key role in assembling the ECM.

Collagen is useful for facilitating tissue regeneration and/or site-specific drug delivery (2) because of its properties such as low antigenicity, low toxicity, high water solubility, and high biodegradability. In particular, type I collagen binds integrins (3,4) facilitating cell migration (5), attachment (4,6), and proliferation and differentiation (3,6). Although atelocollagen, in which the telopeptides on the N- and C-terminals are removed from tropocollagen by pepsin digestion, is widely used in industrial purposes, tropocollagen may be beneficial as the native collagen scaffold containing cross-linked telopeptides.

Chondrex, Inc. provides an acid soluble type I tropocollagen solution which can be used for traditional two-dimensional (2D) systems as well as a scaffold in three dimensional (3D) gels for simulating cell growth in fibroblasts (7,8) and chondrocytes (9). To determine the individual types of collagen, please inquire about Chondrex, Inc.'s "Tips for Collagen Solubilization" protocols, as well as ELISA detection kits for type I collagen and type II collagen. Please visit <u>www.chondrex.com</u> or contact <u>support@chondrex.com</u> for more details

DESCRIPTION:	Acid soluble bovine type I collagen solution		
APPLICATION:	Use for two-dimensional (2D) systems as well as a scaffold in three dimensional (3D) gels for simulating cell growth in fibroblasts and chondrocytes.		
QUANTITY:	4 mg/ml x 12.5 ml (sterile filtered)		
FORM:	Solution in 0.01M HCI		
SOURCE:	Bovine		
ENDOTOXIN:	Less than 10 EU/ml		
PURITY:	>95% Type I collagen		
STORAGE:	4°C		
STABILITY:	6 months		
PROTOCOL:	PLATE COATING (NOTE: an optimized coating condition is required for your culture system)		
	1. Dilute the 4 mg/ml collagen with 0.02M HCl at 50 to 100 μ g/ml.		
	2. Gently mix the diluted solution.		
	3. Add an appropriate volume of diluted collagen solution into wells or plates.		
	NOTE: Ensure the entire surface is coated.		

	4.	Incubate at room temperature or 37°C for 1-2 hours.	
	5.	Remove all solution.	
	6.	Rinse coated surfaces carefully with culture media or PBS.	
	7.	The coated well or plates can be stored at 2-8°C or air dried if sterility is maintained.	
	THREE-DIMENSIONAL (3D) GEL PREPARATION PROCEDURES		
	1.	Dilute the collagen solution with equal volume of cold sterilized PBS (final 2 mg/ml).	
	2.	Add an appropriate volume of the diluted collagen solution in wells or plates.	
	3.	Incubate at 37°C for 30-60 minutes.	
	4.	The gel can be stored at 2-8°C or dried if sterility is maintained.	
NOTES:	N/A		
REFERENCES:			
	1.	M. Shoulders, R. Raines, Collagen structure and stability. <i>Annu Rev Biochem</i> 78 , 929-58 (2009).	
	2.	W. Friess, Collagenbiomaterial for drug delivery. <i>Eur J Pharm Biopharm</i> 45 , 113-36 (1998).	
	3.	Taubenberger, M. Woodruff, H. Bai, D. Muller, D. Hutmacher, The effect of unlocking RGD-motifs in collagen I on pre-osteoblast adhesion and differentiation. <i>Biomaterials</i> 31 , 2827-35 (2010).	
	4.	D. Gullberg, K. Gehlsen, D. Turner, K. Ahlén, L. Zijenah, <i>et al.</i> , Analysis of alpha 1 beta 1, alpha 2 beta 1 and alpha 3 beta 1 integrins in cellcollagen interactions: identification of conformation dependent alpha 1 beta 1 binding sites in collagen type I. <i>EMBO J</i> 11 , 3865-73 (1992).	
	5.	J. Bard, E. Hay, The behavior of fibroblasts from the developing avian cornea. Morphology and movement in situ and in vitro. <i>J Cell Biol</i> 67 , 400-18 (1975).	
	6.	H. Kleinman, R. Klebe, G. Martin, Role of collagenous matrices in the adhesion and growth of cells. <i>The Journal of cell biology</i> 88 , 473-485 (1981).	
	7.	J. Tomasek, E. Hay, Analysis of the role of microfilaments and microtubules in acquisition of bipolarity and elongation of fibroblasts in hydrated collagen gels. <i>J Cell Biol</i> 99 , 536-49 (1984).	
	8.	D. Karamichos, N. Lakshman. W.Petroll, Regulation of corneal fibroblast morphology and collagen reorganization by extracellular matrix mechanical properties. <i>Investigative ophthalmology & visual science</i> 48 , 5030-5037 (2007).	
	9.	S. Oliveira, R. Ringshia, R. Legeros, E. Clark, M. Yost, <i>et al.</i> , An improved collagen scaffold for skeletal regeneration. <i>J Biomed Mater Res A</i> 94 , 371-9 (2010).	

© 2021 Chondrex, Inc. All Rights Reserved, 1202 2.1